In the Specification:

Please amend the specification as shown:

Please delete the paragraph on page 23, lines 10-32 and replace it with the following paragraph:

RNA Isolation and quantitative realtime RT-PCR. Cells were harvested at 2h or 48h after stimulation by scrapping. RNA was isolated and purified using TRIZOL (Life Technologies, Eggenstein, Germany), following the manufacturer's instructions. RNA (2 μg) was reverse transcribed using random hexamers (Boehringer, Mannheim, Germany) and M-MLV reverse transcriptase (Life Technologies, Eggenstein, Germany) in a 50 μl reaction. Expression was quantified with an ABI Prism 7700 Sequence Detection System (Perkin-Elmer Applied Biosystems Inc). Primers were positioned in the coding region and spanned intronic sequences. Internal oligonucleotides (Biotez, Berlin, Germany) were labeled with 6-carboxy-fluorescein (FAM) on the 5' end and 6-carboxytetramethylrhodamine (TAMRA) on the 3' end. Identity of PCR products was verified by sequencing and linearity of each PCR assay were confirmed by serial dilutions of cDNA. Primer pairs and internal oligonucleotides:

rIKCa1: F 5`-CTGAGAGGCAGGCTGTCAATG-3′ **(SEQ ID NO: 27)**; R 5`-ACGTGTTTCTCCGCCTTGTT-3′ **(SEQ ID NO: 28)**; P 5`-AAGATTGTCTGCTTGTGCACCGGAGTC-3′ **(SEQ ID NO: 29)**;

rat myosin heavy chain (rMyHC): F 5′-CATCAATGCCAACCGCAG-3′ **(SEQ ID NO: 19)**; R 5′-TCCCGAGCATCCATTTCTTC-3′ **(SEQ ID NO: 20)**; P 5′-TGAGGCCATGGGCCGTGAGG-3′ **(SEQ ID NO: 30)**;

rat glyceraldehyde-3-phosphate dehydrogenase (rGAPDH): F 5′-CGGCACAGTCAAGGCTGAG-3′ **(SEQ ID NO: 31)**; R 5′-CAGCATCACCCCATTTGATGT-3′ **(SEQ ID NO: 32)**; P 5′-CCCATCACCATCTTCCAGGAGCGA-3′ **(SEQ ID NO: 33)**.

Please delete the paragraph on page 24, line 18 to page 25, line 20 and replace it with the following paragraph:

In situ cell harvesting and reverse transcription. In situ harvesting of single neointimal VSMC from freshly isolated CA segments, isolation of mature VSMC from healthy CA, reverse transcription of mRNA from single cell samples, and "multiplex" single cell RT-PCR performed. First and `nested′ primer pairs spanning intronic sequences for rSlo, small K_{Ca} (rSK1-3), and rIKCa1 were used for the K_{Ca} channels. Primers for rMyHC and endothelial nitric oxide synthase (reNOS) served as markers for VSMC and endothelial cells. Identity of PCR products was verified by sequencing. Forward and reverse primer:

rIKCa1: first: 5′-GAGAGGCAGGCTGTCAATG-3′ **(SEQ ID NO: 1)**; 5′-GGGAGTCCTTCCTTCGAGTG-3′ **(SEQ ID NO: 24)**; nested: 5′-CATCACGTTCCTGACCATTG-3′ **(SEQ ID NO: 2)**; 5′-GTGTTTCTCCGCCTTGTTGA-3′ **(SEQ ID NO: 3)**;

rSlo: first: 5′-GGACTTAGGGGATGGTGGTT-3 **(SEQ ID NO: 5)**′; 5′-GGGATGGAGTGGACAGAGGA-3′ **(SEQ ID NO: 34)**; nested: 5′-TTTACCGGCTGAGAGATGCC-3′ **(SEQ ID NO: 4)**; 5′-TGTGAGGAGTGGGAGGAATGA-3′ **(SEQ ID NO: 6)**;

rSK1: first: 5′-GCACACCTACTGTGGGAAGG-3′ **(SEQ ID NO: 7)**; 5′-AGCTCCGACACCACCTCATA-3′ **(SEQ ID NO: 8)**; nested: 5′-GCTGAGAAACACGTGCACAA-3′ **(SEQ ID NO: 9)**; 5′-TTGGCCTGATCATTCACCTT-3′ **(SEQ ID NO: 10)**;

rSK2: first: 5′-GGAATAATGGGTGCAGGTTG-3′ **(SEQ ID NO: 11)**; 5′-TTTGTTTCCAGGGTGACGAT-3′ **(SEQ ID NO: 12)**; nested: 5′-CTTGGTGGTAGCCGTAGTGG-3′ **(SEQ ID NO: 13)**; 5′-GAATTTCCGTTGATGCTTCC-3′ **(SEQ ID NO: 14)**;

rSK3: first: 5′-AACCCCTCCAGCTCTTCAGT-3′ **(SEQ ID NO: 15)**; 5′-TGTGGTAGGCGATGATCAAA-3′ **(SEQ ID NO: 16)**; nested: 5′-GATAACCATGCCCACCAGAC-3′ **(SEQ ID NO: 17)**; 5′-ATTTCAGGGCCAACGAAAAC-3′ **(SEQ ID NO: 18)**;

rMyHC: first: 5′-CATCAATGCCAACCGCAG-3′ **(SEQ ID NO: 19)**; 5′-TCCCGAGCATCCATTTCTTC-3′ **(SEQ ID NO: 20)**; nested: 5′-AGGCCACTGAGAGCAATGAG-3′ **(SEQ ID NO: 21)**; 5'-TCAATAACTCTACGGCCTCCA-3′ **(SEQ ID NO: 22)**;

reNOS: first: 5′-GAGAGGCAGGCTGTCAATG-3′ **(SEQ ID NO: 23)**; 5′-GGGAGTCCTTCCTTCGAGTG-3′ **(SEQ ID NO: 24)**; nested: 5′-CCAGCTCTGTCCTCAGAAGG-3′ **(SEQ ID NO: 25)**; 5′-ATGGATGAGCCAACTCAAGG-3′ **(SEQ ID NO: 26)**.